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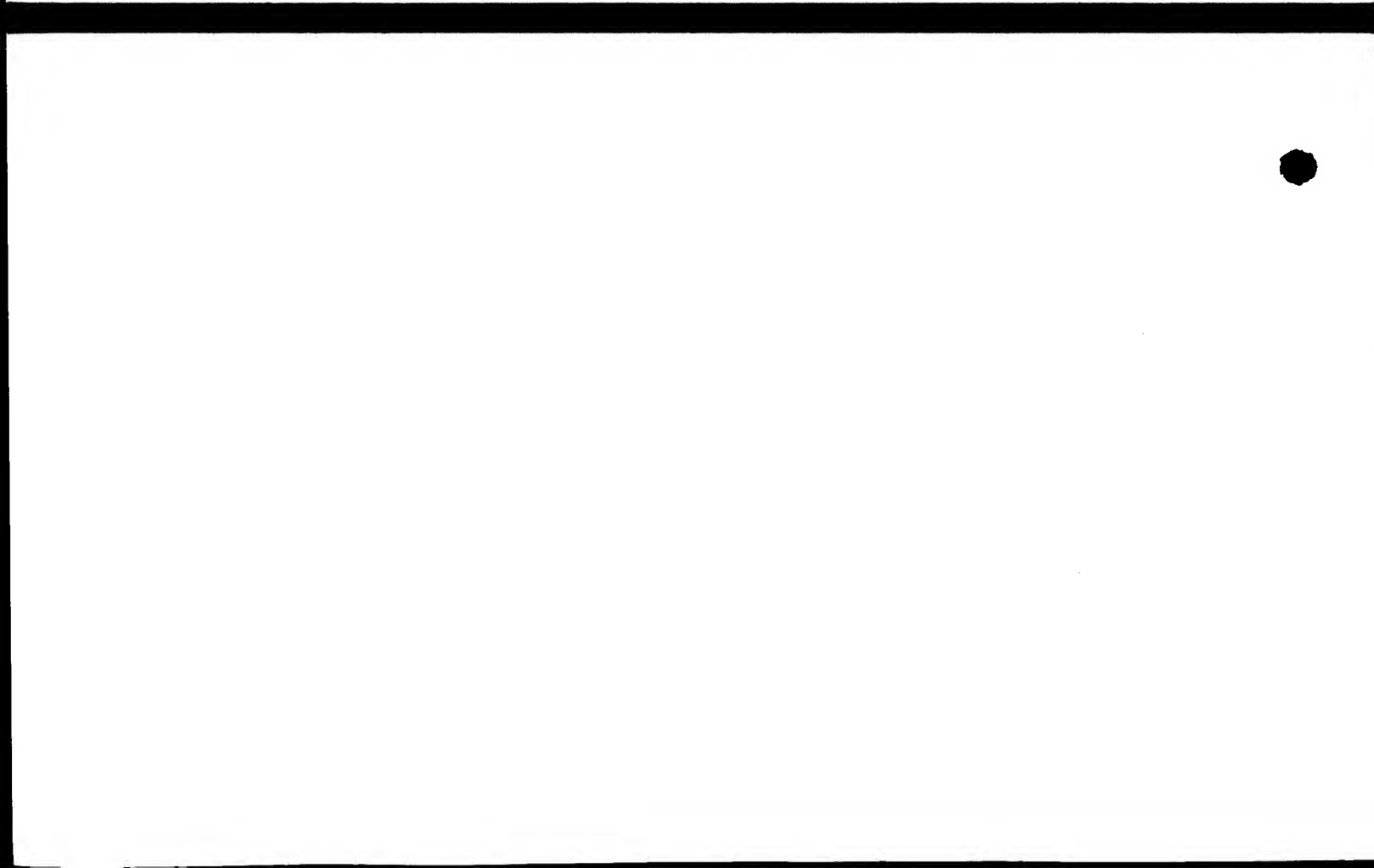
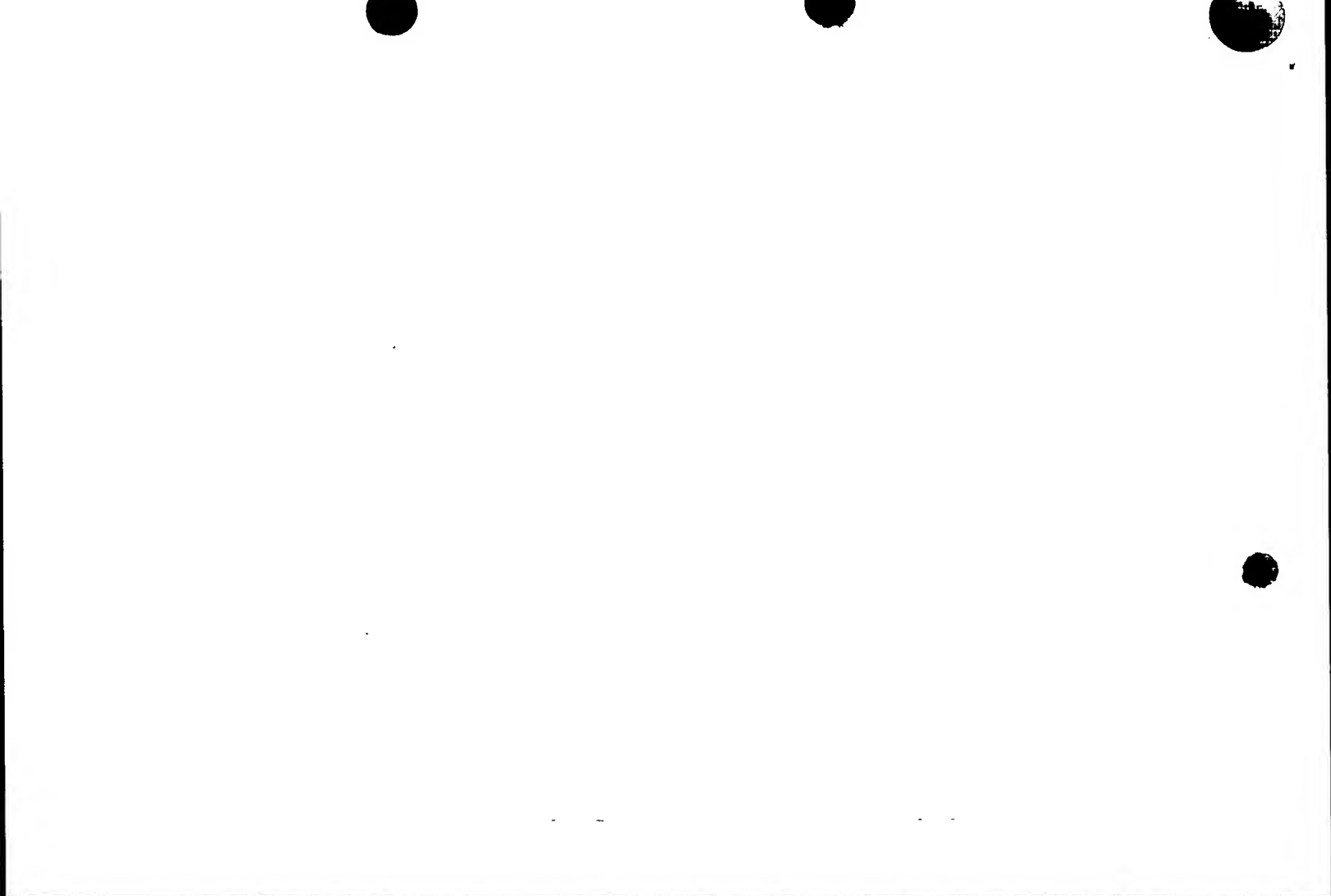
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1. Your reference

C1079.00/C

2. Patent application number

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9914902.3

3. Full name, address and postcode of the or of each applicant (underline all surnames)

Cambridge Imaging Limited
St Johns Innovation Centre
Cowley Road
Cambridge
CB4 4WS

Patents ADP number (if you know it)

If the applicant is a corporate body, give the country/state of its incorporation

06439186001

United Kingdom

4. Title of the invention

Improved Assay Analysis

5. Name of your agent (if you have one)

"Address for service" in the United Kingdom to which all correspondence should be sent (including the postcode)

Keith W Nash & Co
90-92 Regent Street
Cambridge
CB2 1DP

Patents ADP number (if you know it)

1206001

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Country

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Number of earlier application

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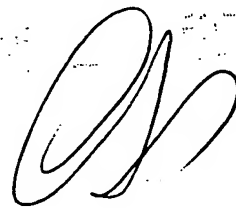
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Description 23

Claim(s)

Abstract

Drawing(s) 6 + 6



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KEITH W NASH & CO, AGENTS

12. Name and daytime telephone number of person to contact in the United Kingdom

Mr K W Nash (01223) 355477

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C1079/C

Title: Improved assay analysis

Field of invention

This invention relates to the measurement of radiation, typically fluorescence emitted by samples and assays in the field of biological, bio-medical and chemical sciences.

Background to the invention

Such assays are normally prepared and measured in sample plates or formats, including 96-well microtitre plates, petri dishes, gel matrices, membranes, glass slides and capillaries. The trend is towards higher throughput detection of samples and the use of smaller volumes in each of the samples, resulting in so-called miniaturised sample formats.

This requires the corresponding development of detectors capable of handling such miniaturised formats. This is particularly so in the case of high throughput screening (HTS) of biological assays as applied to drug discovery and the screening of drug candidates.

Such miniaturisation is achieved by arranging samples for assay and detection in well plates in which typically there can be 96, 384, 864, 1536 or 3456 wells per plate, and sample volumes can vary from 200 microlitres to as little as 1 microlitre.

Alternative formats for the miniaturisation of assays include capillaries, microchannels or microfluidic structures, including microwells, which can be moulded or etched in substrates such as glass (eg silica or quartz) or plastic. In these alternative formats, the sample volumes can be of the order of nanolitres and picolitres.

In order to achieve high throughput screening it is necessary to interrogate large numbers of such samples simultaneously. In the case of fluorescence based assays, detection or interrogation consists of illuminating each sample with excitation light, and subsequently detecting the emitted fluorescence from each sample separately. Examples of fluorescent based processes include prompt fluorescence and time-resolved fluorescence where there is a time delay between photoactivation of the sample and emission, in the range, for example of ps to ms or more. A further process is fluorescence or luminescence energy transfer. In this process a molecule is activated, for example by excitation light, and transfers energy via eg resonance energy transfer or chemical transfer to a second molecule, which in turn emits light. This process can involve different or multiple secondary molecules, which can emit radiation over a range of wavelengths. Further examples of luminescent processes include phosphorescence, and chemi- and bio-luminescence. Wavelength ranges for all these processes include UV, visible, red and infra-red (approx 250-1200nm).

In a typical arrangement, a scanning head with 96-channels simultaneously interrogates the 96-sites arranged in the 8 x 12 pattern of a 96-well microtitre plate. By stepping a larger-well density plate relative to the 96-channel scanning head, the remaining sites can be read eg 2 x 2 steps will cover 384-samples, 6 x 6 steps will cover 3456 samples, and so on, so as to address a high density sample presentation format.

Fluid samples, eg liquids or gels, can be placed in small sample sites, such as micro-capillaries or microchannels, which can be typically 100 x 100 um in section, and typically 1-100mm long. The samples can be moved by pumps, or electrophoretically or electro-osmotically. Samples can be solid or a matrix, such as beads, agarose or microparticles, suspended or otherwise contained in a fluid medium, including a gel type format. Other samples can comprises suspensions or

monolayers of cells.

Applications include cell biology, hybridisation techniques and immunoassays including binding assays. In such assays, materials can be labelled with a fluorescence marker for the purpose of identification. In a binding assay, a bound molecule can be separated from an unbound molecule, as between a solid and liquid medium. Further applications include electrophoresis or electro-osmosis fluids such as liquids in gels and media including agarose. Such applications can be run in miniaturised formats including micro-capillaries and micro-fluidic structures, where molecules or moities may be separated spatially by properties including molecular weight or charge and may also be labelled with fluorescent or luminescent tags or dyes for the purpose of detection and identification. The techniques described herein can be applied to the detection of biological compounds including proteins and nucleic acids, and in cell biology, processes such as cell signalling or cell binding can also be detected.

Separated molecules can be contained in a fixed matrix, such as a gel or agarose beads, or can be separated in a fluid such that the separated molecules or moities will flow at different rates through a medium and can be detected at a fixed point along the flow path as a series of emission peaks based on their time separation profile. It is important that emission detection methods posses high accuracy and sensitivity for the determination of such peaks, and the rapid and/or continuous and/or simultaneous measurement of samples.

A number of such assay techniques involve time changes in light emission. To measure such changes requires an ability to perform rapid, accurate, sensitive and repetitive readings, that is to perform kinetic measurements. In order to achieve high throughput it is necessary for a system to measure multiple samples simultaneously, requiring high resolution, high sensitivity and high signal detection efficiency.

Mostly the samples/assays are arranged to emit light in the middle range, although the quantities of light per sample can be very low.

A further issue with fluorescence-based assays is the problem of quench which causes a reduction in light emission. This can occur in samples and assays particularly those involving cells, due to chemical effects which interfere with the light signal, or due to coloured substances or particles in the sample or medium, which reduce the light signal. In certain applications, such as inhibition assays, a change or reduction in light signal is the feature of the assay requiring measurement.

Figure 1(a) shows an array of 8 x 12 micro capillaries in a substrate, in which either a liquid, or molecules or moities in a fluid, move in the X-direction. This is an example of an array of miniaturised samples such as will be referred to in this application.

involve high density or miniaturised sample formats..

Background to the invention

A confocal microscope is a means for achieving excellent imaging capability whilst possessing a unique depth-discriminating property. (A typical Zeiss system is shown in Figure 15.5 in "Quantitative fluorescence microscopy", F.W.D. Rost, CUP 1991). A parallel light beam from a laser or other suitable light source can be used for fluorescence excitation. The beam is focused by a microscope objective, down to a spot of typically micron or submicron size, within the sample. The fluorescent light emanating from the spot is focused by the same objective, via a beam splitter, onto a pinhole aperture (a spatial filter), and thence onto the detector. The detector does not accept all the light from out-of-focus planes and so

these are imaged less strongly by the spatial filter. Thus detected background fluorescence from the substrate or material surrounding the sample, which is also unavoidingly illuminated, is sharply reduced relative to the wanted signal. Moving the microscope relative to the plurality of sample spots or vice versa, allows the spots to be inspected in sequence or scanned.

A confocal microscope therefore has the essential attributes for interrogating the tiny samples described above, such as micro capillaries.

Firstly, there is excellent spatial resolution to excite a small region of a sample and avoid exciting laterally surrounding material.

Secondly, it has good depth discrimination, which minimises the effects of illumination of the substrate, above and below the sample. Hence the effect of background fluorescence at the detector can be reduced/minimised.

Single channel scanners for the purpose have been reported in the literature. For example, we refer to "DNA sequencing using Capillary array Electrophoresis" by X C Huang, M A Quesada and R A Mathies, Anal. Chem. 1992, 64, 2149-2154. In their apparatus a 32X. NA 0.4, Carl Zeiss microscope objective brings the parallel beam from a 1mW argon ion laser to a 10 micron diameter spot, within a 100 micron id sample capillary. The fluorescence is collected by the same objective, passed via a beam splitter and, after wavelength selection via spectral filters, is focused onto a 400 micron diameter pinhole. The spatially filtered light is then detected by a photomultiplier. In this apparatus a set of capillaries mounted side by side is interrogated successively by the confocal microscope, by means of relative XY movement between the samples and the microscope. However a serious disadvantage of this approach is that no sample is read continuously but is read at best, on a periodic basis. Typically any one sample can be read about once per

second.

Thus a single channel confocal scanner has been used to measure a number of sample sites but only sequentially and not simultaneously, and such a system is not appropriate for high throughput scanning where simultaneous detection and assessment of a large number of sites is required such as in a 96 well microtitre plate or higher density format array or array or microcapillaries.

Summary of the invention

According to one aspect of the present invention, a plurality (N) of single channel confocal optical systems and photoelectric detectors are arranged in parallel to form a plurality of reading heads arranged side-by-side so as simultaneously to read a corresponding plurality of adjacent sites.

The sites may be in a column or in an array such as a

The optical systems may be arranged in a single line for reading column by column of a multi-column array, or in a staggered pattern for simultaneously viewing sites in different columns if this were preferred.

This system would comprise essentially (N) independent confocal systems, each with its optic axis aligned with one sample site.

Advantageously only one laser source is provided which may be split into an appropriate plurality of beams, conveyed each by a fibre optic cable to individual sample sites.

The light emitted from the separate sites may be conveyed to individual detectors, or discrete regions of an array detector, via optical fibres.

Preferably the reading heads are independently adjustable so each can be positioned accurately over the samples. To this end a special opto-mechanical device may be provided to bring each scanner's optic axis into alignment by means of an appropriate scan in the Y-direction.

Where there is a tolerance in the position of the samples, such as in the case of capillaries, which result in uncertainty in the Y-direction, adjustment is preferably provided.

Such an arrangement could be used to inspect say 8 or 12 sites at a time, but an arrangement to inspect larger numbers simultaneously could be costly, complex, and cumbersome. We would refer to such arrangements as "ganged" systems.

According to a more preferred aspect of the invention in a method of imaging a plurality of micro-sample light emitting sites simultaneously onto separately addressable detectors, so that light emitted from each site can be monitored by one of the detectors, a corresponding plurality of objective lenses are located adjacent to the micro-sample array with one objective lens for each micro-sample, the latter are located at or near the focal point of each of the lenses so that light emanating from each micro-sample is collected by its respective objective lens and converted into a beam of parallel or near parallel rays, the objective lenses are arranged so that the axes of all the beams issuing therefrom are parallel and spaced apart, the beams are focused by a focusing lens through a single point and collected beyond that point by detector lens means which serves to reconstitute the parallel beams for presentation to the detector array.

If near parallel the rays may be converging or diverging and the optical system downstream from the objective is adapted accordingly.

The axes of the objectives may be angled if desired with

appropriate adjustment of the optical characteristics of the objectives and/or downstream optical system being adapted accordingly.

Preferably and advantageously a single focusing lens (which may be a multi-component lens) is employed for directing all the beam paths through the single point.

The apparatus can be thought of as a single channel imaging system having a plurality of separate paths therethrough the separation of light between one path and another being controlled by careful choice of the quality of the optical components forming the various lens assemblies.

In general the light emitted from the micro-samples will comprise fluorescence of components of the sample. Where the fluorescence is initiated by excitation illumination, fluorescence may be triggered not only from the specific component of the sample which is to be monitored, but also from material around and upstream and downstream of the region of

emanating from upstream and downstream regions can be improved by placing a small aperture pinhole at the focal point of the focusing lens means so that light which is not emanating from the focal point of each of the objective lenses adjacent the micro-sample sites will be out of focus at the small aperture pinhole.

If the aperture size is appropriately selected, out of focus light will be significantly attenuated. On the other hand light emanating from a point in a micro-sample at the focal point of its micro-sample objective lens, will pass through the pinhole relatively unattenuated.

By enlarging the pinhole aperture, more (possibly unwanted) background originating light may pass therethrough but this may be acceptable if in so doing more light can be focused onto the

detector from a larger volume within the sample.

According to a preferred feature of this aspect of the invention, the micro-samples are positioned relative to the micro-sample objective lenses so that the region of interest is as close as possible to the focal point of the respective objective lens.

For simplicity, the samples are located on a planar support with the regions of interest all in the same plane so that the objective lenses can likewise all be in the same plane parallel to that containing the regions of interest in the samples.

The method includes the step of adjusting the position of the micro-sample array relative to the lens array and also includes the step of individually adjusting the position of at least the objective lenses relative to the micro-samples or vice versa to ensure that the regions of interest in the micro-samples are at the focal point of the respective micro-sample objective lenses.

Where spectral separation is required based on wavelength, a suitable filter can be included in the light path either between the micro-sample objective lenses and the focusing lens means ahead of the pinhole, or between the detector lens and the detector array.

Preferably any such filter is located in a region in which the light paths are parallel or nearly parallel.

Where fluorescence is the mechanism which generates the light which is to be focused onto a detector, excitation radiation to produce the fluorescence is advantageously applied only to the region of interest within each micro-sample rather than over the whole of the micro-sample. To this end the optical elements which determine the path to be traversed by light emanating from the region of interest in each micro-sample, can

be used to advantage to inject the excitation radiation onto the micro-sample region of interest, since by doing so, the excitation radiation can be focused onto the region of interest rather than broadly illuminating more or less the whole of the micro-sample.

To this end according to a further preferred feature of the invention, excitation radiation is injected into the multipath optical system so as to proceed in a parallel sense towards the array of objective lenses, in an opposite sense to the light which emanates from the micro-samples, so as to be focused by the objective lenses onto the region of interest in each micro-sample.

Typically the excitation radiation is injected as a parallel beam into the optical path, at right angles thereto, and a 45° beam splitting device is provided onto which the parallel excitation radiation impinges and from which it is directed in a parallel manner towards the micro-sample imaging lenses, but through which light from the micro-samples can pass to the

Typically excitation radiation is produced using a laser such as an argon ion laser, and a beam expander is employed to expand the cross-section of the laser beam into a relatively large area beam equivalent to the area of the parallel array of micro-sample objective lenses.

Where optical light is employed, the beam splitter may be a dichroic mirror.

Whilst in theory the invention can be employed for any number of micro-sample objective lenses, it is advantageous to arrange an 8 x 12 array of micro-sample objective lenses on the same 8 x 12 matrix as a standard 96 well plate, and if the imaging system is to be used to inspect for example a 384 well plate, the latter is mounted on an XY stage so that it can be moved

relative to the array of objective lenses in manner known per se so as to present groups of 96 wells making up the 384 wells, to the 96 lens array. Thus a 2 x 2 movement of the XY stage would be required to complete the inspection of all of the well sites in the 384 well array.

According to another preferred feature of the invention, the parallel beams of light directed towards the detector array may be transferred to the latter via optical fibres, preferably in the form of a fibre optic bundle or fibre optic plate.

According to a further preferred feature of the invention, where a bundle of fibres is employed, the arrangement of the fibres in the bundle may differ between the input and output ends thereof so that the array of fibres which convey the light paths to the different regions of the detector array conforms more to the shape of the detector array in the XY plane and enables for example, optimal utilisation of a generally square array by the rectangular proportions of an 8 x 12 micro-sample array.

The detector array is typically a charge coupled device having a large number of separately addressable regions (each of which is commonly referred to as a pixel) and groups of adjacent pixels (or individual pixels) from the detector for each sample, to enable good resolution to be obtained in the XY sense as between the light from one sample and another.

The charge coupled device may be cooled, for example cryogenically so as to reduce background noise signals associated therewith.

Additionally or alternatively an intensified CCD device may be employed so as to increase the sensitivity of the detector to the relatively low light levels typical of bioluminescent assays.

The invention is not limited to the nature of the detector or to the technique in which the different regions of the detector are addressed to enable light emanating from one sample to be distinguished from light emanating from another, nor is the invention limited to any of the circuits or computing techniques which may be employed for processing electrical signals obtained by addressing the different regions of the detector array.

Further lenses for focusing the parallel beams of light directed towards the detector array may be employed, for improving the resolution at the detector surface either in combination with a fibre optic transfer bundle, or otherwise.

Since the spacing between, and the actual size of, the sample sites in a micro-sample array, are very small, and can be of the order of microns, micro lenses, optionally in combination with a fibre optic transfer plate may be employed in the objective lenses adjacent the micro-samples.

microscope objectives in that they are short focal length lenses designed to form an image of a tiny object (in this case light being emitted from a small region in a micro-sample). However as distinct from a conventional microscope objective, the optical characteristic of the sample objectives is such as to produce a parallel beam of light therefrom so that in this sense the objective can be said to have one infinite conjugate.

By utilising high quality components and projecting the excitation radiation to the samples via the objective lenses, the excitation radiation can be focused to precisely the regions of interest in the samples, to reduce excitation of surrounding sample material.

Where a filter is located ahead of the detector, apertured masks may be placed on either side of the filter to collimate

the parallel beam to further reduce background and cross-talk.

The invention also lies in apparatus adapted to perform the above methods and comprising means for supporting a micro-sample array on a substrate in close proximity but parallel to an array of micro lenses arranged so as to correspond on a one to one basis with the positions and spacing of at least some of the micro-samples on the substrate, each of the micro lenses being positioned relative to a region of its related micro-sample by a distance equal to the focal length of the lens so that light emitted from that region of the micro-sample will emerge from the lens as a parallel beam parallel to the axis of the lens, and the parallel beams of light are focused by means of a single focusing lens onto a detector lens so as to produce an image of the micro-sample light emissions in the plane of an array of individually addressable photoelectric detectors, such as regions of an addressable CCD array, and circuit means is provided to which signals read out from the array are supplied in the form of a sequence of digital values or otherwise, each corresponding to the light incident on a region of the detector for a given period of time from one of the micro-samples, and computing and analysing circuit means is provided, responsive to the electrical signals, together with memory means for storing data indicative of the light found to be emitted from each of the micro-samples, for storing those values together with address information, whereby each stored value can be identified with the micro-sample on the substrate from which the light giving that value has been emitted by reference to the position of the region in the detector array and by correlating the position of the sample in the sample array.

The apparatus may further include a beam splitter, such as a dichroic mirror, interposed in the optical path between the micro lenses and the focusing lens to enable on the one hand light to pass from the lenses to the focusing lens, and on the other hand to enable excitation radiation, typically light of

a particular wavelength, to be reflected as a parallel beam towards the micro lenses, thereby utilising the optical focusing characteristics of the micro lenses to focus the parallel light into spots of light which register with the micro-samples so that the latter are individually radiated by excitation light which is predominantly incident on that region of each micro-sample which is to be inspected for fluorescence after the excitation radiation has been removed, and filter means may be provided in the optical path between the beam splitter and the detector array to generally attenuate any excitation wavelength radiation travelling towards the detector and generally prevent such radiation from reaching the detector.

The apparatus preferably includes a pinhole aperture at the focal point of the focusing lens so as to improve the on-axis resolution of the optical system and assist in attenuating unwanted fluorescence such as from background material and other components of an assay ahead of or behind the region of interest in a micro-sample measured along the optical axis of the objective, from reaching the detector.

The apparatus typically includes a laser source, as the source of excitation radiation a beam expander for enlarging the cross-section of the laser beam and presenting a generally uniform parallel beam of excitation radiation for entry into the imaging system via the beam splitter or dichroic mirror.

Shutter means may be provided to inhibit the passage of the laser light, except when required for excitation purposes, and further shutter means may be provided synchronised with that associated with the laser source to prevent light of any wavelength reaching the detector whilst excitation light is projected into the system.

The invention also comprises a method of analysing fluorescence emitted by radiation excited samples in an array of samples

comprising the steps of focusing light emitted from each said sample at infinity so as to form a parallel beam, in parallel with the light from all of the other sample sites making up the array, subsequently focusing all the parallel light paths through a single point and locating at the point a small aperture to restrict unwanted light from fluorescing material upstream and downstream of the sites of interest in the samples, and re-establishing a parallel array of light beams by the use of a further lens so as to present to an addressable detector array a plurality of parallel light paths corresponding to the light paths from the samples, and individually addressing different regions of the detector array onto which the parallel light paths impinge, to determine the light incident thereon, and storing data relating to the quantity of incident light on each said region of the detector array together with address information to enable the data to be reconciled with the position of the sample in the array on the substrate to which that data relates.

The method typically further comprises the step of introducing periodically into the optical system excitation wavelength illumination and projecting same through the optical imaging devices associated with the array of samples to project the excitation illumination onto a specific region in each said sample, thereafter extinguishing the excitation wavelength light and enabling fluorescence caused by the excitation to pass through the same optical devices to emerge as parallel rays of light for transfer to a detector, for analysis as above mentioned.

The objectives may be arranged above or below a sample array.

The method and apparatus described herein may be used to perform immediate fluorescence analysis (as so far described) or time resolved fluorescence analysis in which a shutter is provided to inhibit the transfer of light to the detector, excitation radiation is supplied for a short interval of time

and then sent off (either by pulsing the source out using further shutter or both), after a selected interval of time the shutter preventing transfer of light to the detector is opened and after an appropriate integration interval (which can be very short or longer as described) the residual charge pattern on the charge coupled detector array is interrogated to generate a signal relating to the charge pattern (and therefore indicative of light incident thereon) for processing and storage as aforesaid.

The invention will now be described, by way of example, with reference to the accompanying drawings, in which:

Figures 1A, 1B and 1C illustrate arrangements of sample arrays as previously mentioned;

Figure 2 is a schematic diagram of a confocal imager for measurement of micro-sample assays such as may be contained in an 8 x 12, 96 well microtitre plate;

viewing lenses in relation to the samples and typical scan range across each sample;

Figure 4 is a ray diagram indicating in detail the optical arrangement of the pinhole aperture which improves the on-axis resolution of the system; and

Figure 5 is a further schematic and ray diagram illustrating how an 8 x 12 array of light spots can be re-arranged into a pattern having a generally square outline which is more applicable to a typical CCD camera target.

Figures 2 onwards illustrate a unitary, optical scanner which can measure a large number, say 96, of channels simultaneously. The system shown is essentially 96 confocal scanners in parallel, but having a single optic axis, and particularly a

single spatial filter in the form of a pinhole aperture. It has all the advantages of scanning confocal microscopy, such as high spatial resolution and good depth discrimination, to interrogate selectively the array of 96 samples, whilst tending to reject fluorescence background from the substrate and surrounds of each of the 96 sites.

One multi channel confocal scanner arrangement is shown in Figure 2. The example assumes 96 channels.

A laser with sufficient power to illuminate 96 sites simultaneously, comprises a 1 W argon ion laser, followed by a beam expander of suitable diameter and a beam splitter. This can deliver at least 10 mW of excitation light (eg of 485nm for fluorescein) to each of 96 suitable lenses arranged with parallel optic axes in an 8 x 12 pattern under an XY stage supporting a multiple micro site assay such as a 96 well microtitre plate.

Alternatively, one could use a conventional, eg filament or arc, source with lenses and with spectral filters to achieve the broad parallel beam required wavelength.

Each lens creates a spot of illumination of ~ microns in diameter in the region of its micro-site. A suitable lens would be a microscope objective of 10X with NA 0.4, with focal length about 8mm, and aperture diameter 5mm.

A normal microscope objective is designed to produce an image of a tiny object being examined at an intermediate focus. This intermediate focus is in turn viewed through a eyepiece. In this invention however, when using a conventional microscope objective, it is preferable for this intermediate focus to be removed to infinity, eg by using a negative (concave) lens (not shown) beyond the microscope objective. In this way, incoming parallel excitation light will produce the required focal spot of excitation light, and a source of fluorescent emission light

at the focal spot will likewise produce a beam of parallel light directed towards the focusing lens (see Figure 2). Another way of putting this is to say that a conventional microscope objective is modified to give optimum spatial resolution with one infinite conjugate.

The emitted fluorescent light returns parallel to the axis of each microscope objective, so that the 96 samples result in a wide parallel beam which then passes through the beam splitter into a focusing lens, which focuses the light through a single pinhole aperture (spatial filter). This simultaneously allows through all 96 signals but tends to reject the background fluorescent light in any of the channels. This rejection is achieved by the depth discrimination capability of confocal microscopy, since this background fluorescence arises from substrate which is in front of or behind each sample spot and so is out of focus at the pinhole. Also the spatial resolution (say about 10 microns) is sufficiently good to select out background fluorescence coming from material surrounding laterally each fluorescence sample.

The pinhole diameter may be adjusted to optimise the depth and lateral acceptance. This could be used to adjust the amount of signal measured and the proportion of background that is accepted.

Finally, the light diverging from the pinhole is received by the detector lens of Figure 2, whose focal plane is at the pinhole. This lens produces a parallel beam which can pass through one or more spectral filters, as required.

At this stage the light from each of the 96 samples will form an array of 8 x 12 bright regions, with separation from each other determined by the separation of the 96 microscope objectives and the focal lengths of the focusing lens and the detector lens. Each bright region will be of diameter approximately equal to that of the microscope objective, but

broadened by any residual lens defects and edge scattering at the pinhole, and then reduced by the ratio of the focal length of the detector lens to the focal length of the focusing lens. A detail of the final part of the system, namely the detector optical arrangement, is shown in Figure 4 which illustrates the 96 parallel beams of light passing through the filter. A mask may be placed on either side of the filter, as shown, to collimate each of the beams as a means of further reducing background (due to scattering and optical defects), and cross-talk.

This array of 8 x 12 bright regions is detected by a suitable imaging array, for example an interline CCD with say, 10 Hz frame rate, and of sufficiently small pixel size to give the required spatial resolution. With 10 Hz frame rate one can obtain a succession of images of almost 0.1 second duration, after allowing for dead-time in the detector. Alternatively, an image intensifier or an image intensifier coupled to a CCD, or a series of array of photodiodes, or PMT's, may be used as the detection means. Cooled versions of these detectors may be used, to reduce noise.

To produce an image of the 96 bright regions one could optionally insert an array of 96 micro lenses as shown in Figures 2 and 4, position so as to image each region down onto a spot of small size (the order of a millimetre) at the surface of the detector. This will give better definition of each spot on the detector, allow a lower spatial resolution detector to be used, and concentrate the signal over a small area of the detector, hence reducing the effect of background noise in the detector.

Figure 5 shows an alternative detector optical arrangement to that shown in Figure 4, in which an array of fibre optic cables is located immediately after the filter, to gather the light from each of the 96 parallel beams emanating from the filter. The far ends of the fibres may be mapped into any arrangement

to match the aspect ratio and/or shape and/or area of the detector.

This design is less complex and costly than a "ganged" system of 8 or more independent confocal scanners. It could be extended to more than 96-channels and could also be scaled in size, to examine smaller arrays of tiny sites on an area much smaller than a microtitre plate, for example this could be a high density plate, such as one containing 3456 samples, or a gridded array on a membrane or gel. This could be achieved using smaller components as appropriate, eg an array of micro lenses instead of microscope objectives. To achieve a good result it will obviously be necessary to use high quality lenses throughout, to have the optic axes of the lenses in the scanning array accurately parallel, and to have minimum light scatter from edges etc. This is particularly important for the design of the pinhole aperture, to avoid scatter which could lead to cross-talk between channels.

Another advantage of this invention is that all sites are

time for the whole array means a longer dwell time on a given site and hence more signal and better measurement accuracy.

Note that in Figure 2 the illumination system, i.e. laser plus beam expander, and the detection system, i.e. focusing lens pinhole, optics and detector, could be interchanged with respect to the beam splitter. The properties of the beam splitter, namely the respective wavelength range of reflected light and transmitted light, would be chosen accordingly. Examples of such a wavelength selecting device include a dichroic mirror or band pass filter. Likewise optical reflectors or mirrors may be used in conjunction with this overall arrangement, to fold, contact or otherwise rearrange optical paths, without affecting the system performance.

In Figure 3 part of a 96-site array is shown positioned

relative to the corresponding microscope objectives. In this example each site comprises a microcapillary aligned in the X-direction, having a width of say, 100 μm , in the Y-direction. The depth of the capillary (in the Z-direction) would be typically 50-100 microns. Because of the tolerance in the Y-position of each capillary mentioned above, it would be necessary to move the XY stage in a number of steps in the Y-direction, say 10 steps of 20 μm size. At each step a reading, say of 0.1 second duration, is taken. About one-half of the readings will be taken when the scanning spot is within the capillary, and so give useful signal data. The balance of readings would be predominantly of background while the scanning spot moves outside the capillary, and these could be useful as they would give more information about and monitor background in case a correction has to be made for background in the signal.

It should be noted that if all the samples are large enough, or sufficiently well defined in location, such as in the Y-direction in our example in Figure 1(a), then scanning might not be necessary. The axes or spots of the multi-channel confocal scanner can be located appropriately on the multiple samples, and measurements made of all samples simultaneously. High density formats can be read by a series of steps across the matrix, as previously described..

In this example 10 readings of the sample are acquired on a continuous basis per second, for each of the 96-sites simultaneously. This rate can be increased as necessary using a faster read-out detector.

This ability to continuously monitor all the sample sites simultaneously has the following advantages in comparison with existing systems which sample periodically, or only sample a small number of the sites at any time.

Firstly the method allows a longer dwell time per site per

reading, and hence allows more signal to be detected and hence improves sensitively and accuracy.

In the case, for example, of samples flowing past the detector, such as in micro-capillary electrophoresis where separate peaks pass the detector, the continuous monitoring capability allows better definition of the sample peaks and also allows the flow rate to be monitored and measured more accurately.

Likewise where the level of light emission is changing with time, such kinetic features can be monitored more accurately.

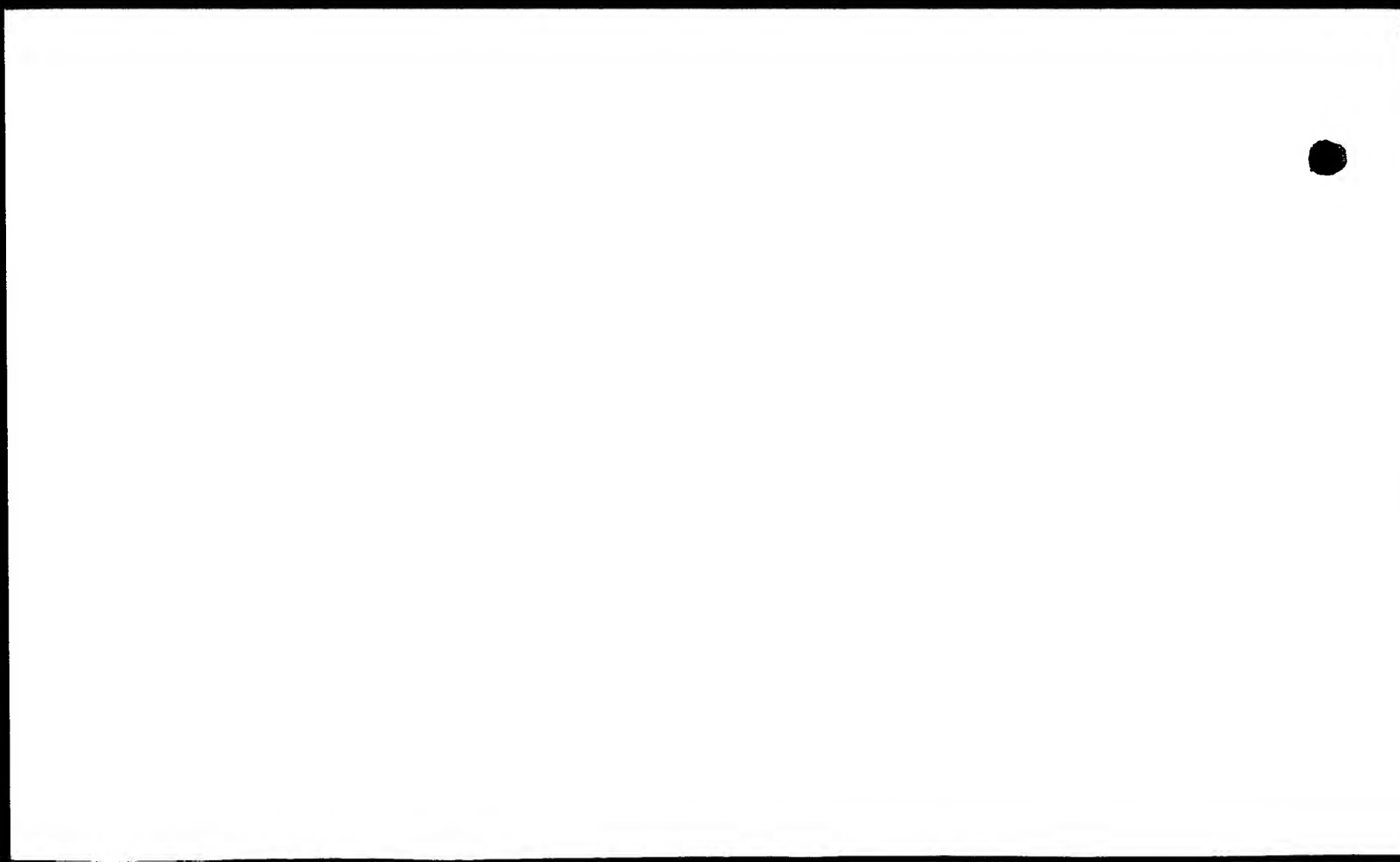
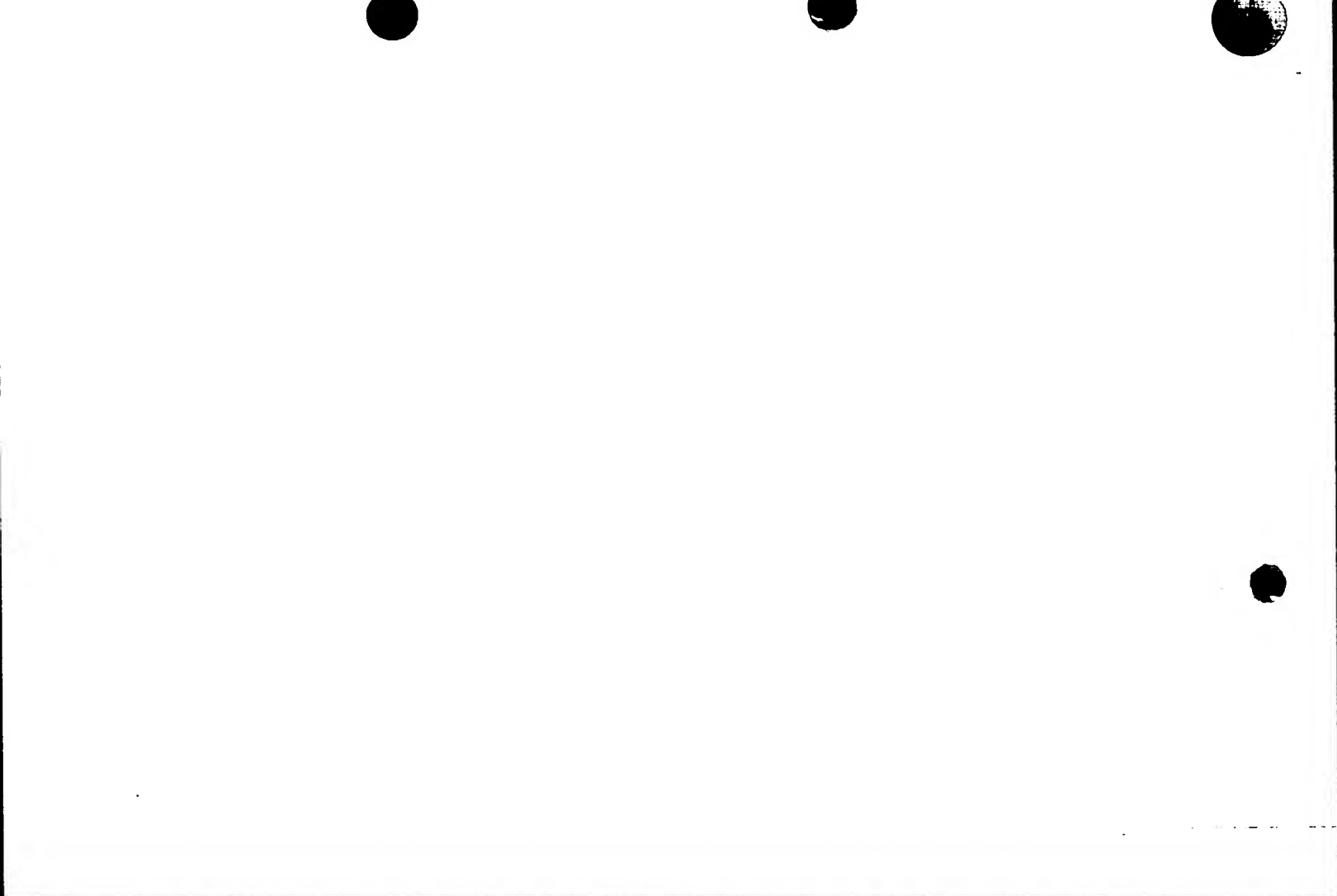
The continuous monitoring capability also allows any quench effect of the light signal, i.e. the interference with or the reduction of the light emission from the sample, to be measured.

In separation processes such as electrophoresis, the molecules or moities in the sample are caused to move in a particular direction (e.g. the X-direction in our Figure 1(a)), and this

measured by a detector channel as the peaks pass along the fluid matrix. The speed of motion can be chosen within limits. One could take a series of, say 50, sets of readings, each set taking 1 second, so that the plate would take 50 seconds for complete measurement. Assuming a given peak passes the reading spot in 5-10 seconds, this means that one will obtain 5-10 sets of readings of the given peak, each set giving about 5 useful readings of 0.1 seconds each. This ought to be sufficient to give an accurate estimate measurement of the total fluorescence signal from a peak. Several such peaks could be interrogated in the 50 seconds indicated here for the whole measurement process of a plate.

The invention is not limited to the use of any particular fluorophore or wavelength, nor to the specific optical arrangements illustrated herein. Thus the invention is

applicable to a wide range of fluorophores and wavelengths and variations can be made in the optical systems described, without departing from the scope of the invention.



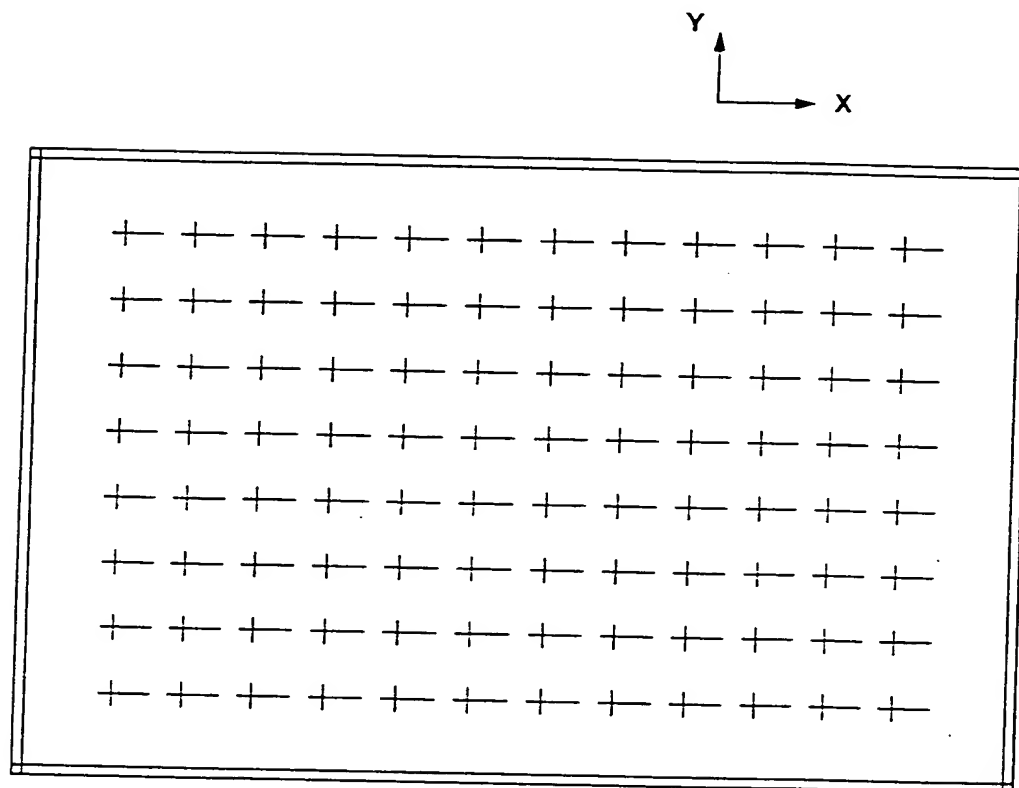
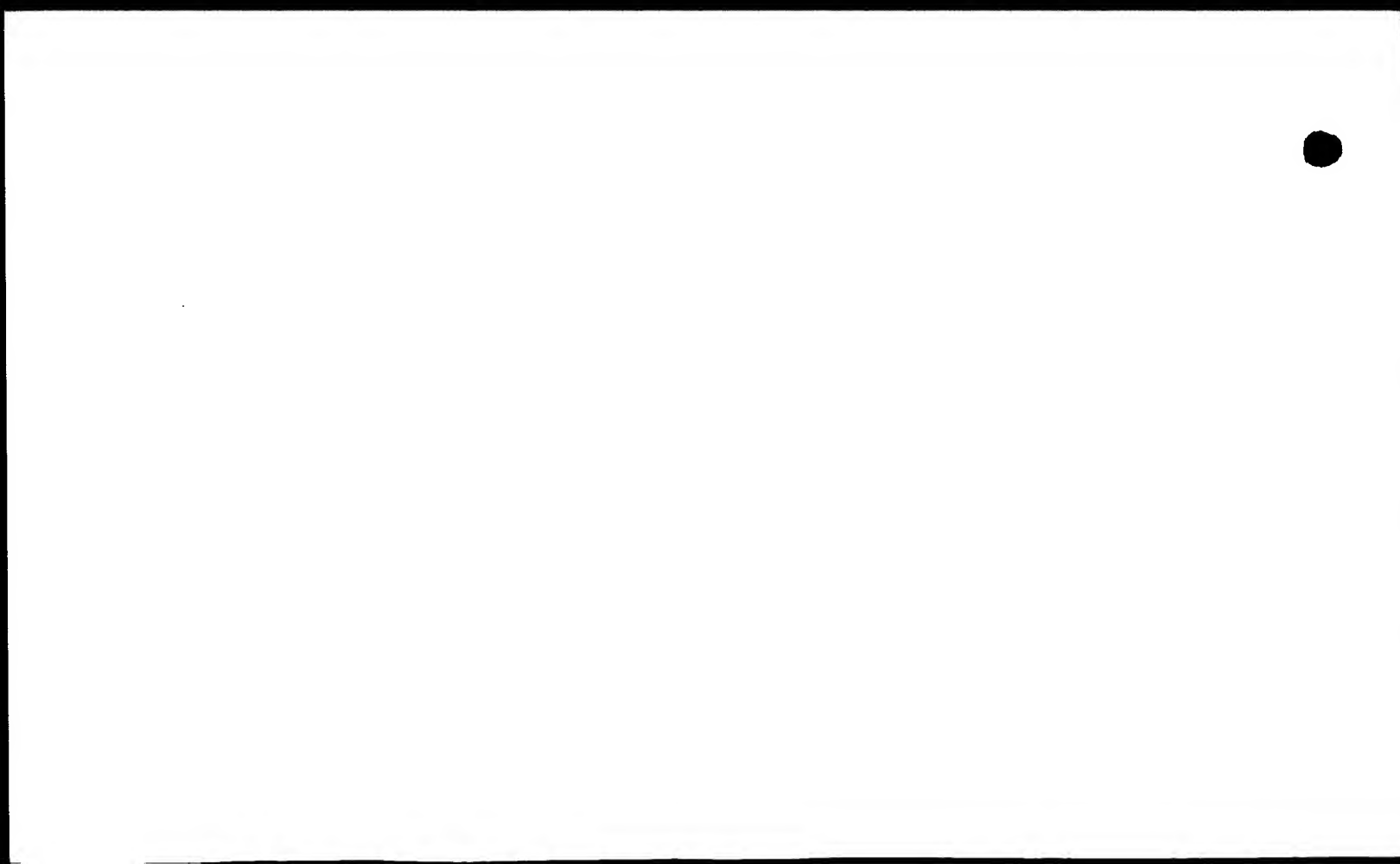
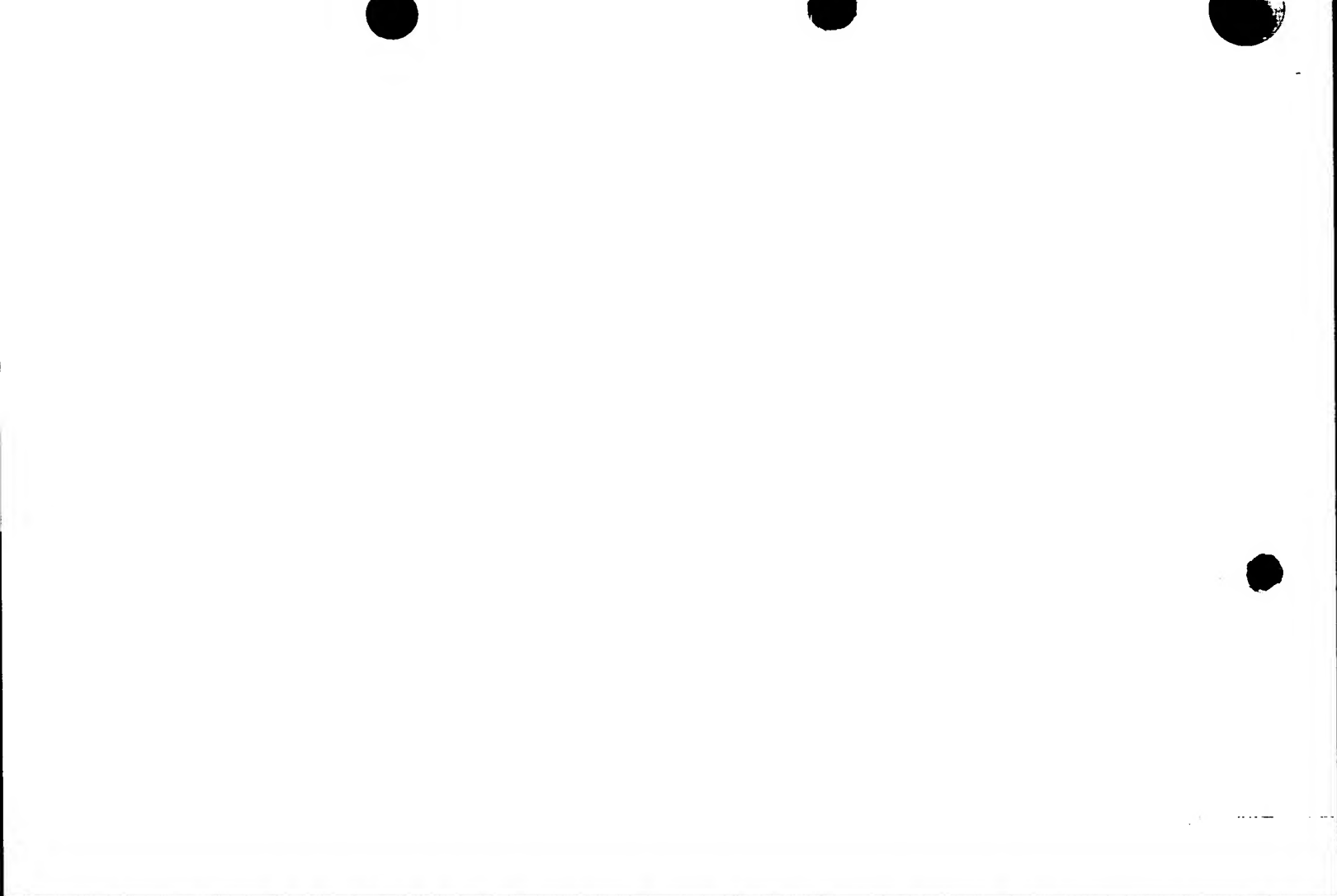


Fig 1(a).

Example of microcapillary channels arranged in an 8 x 12 pattern, similar to the wells in a 96-well microtitre plate



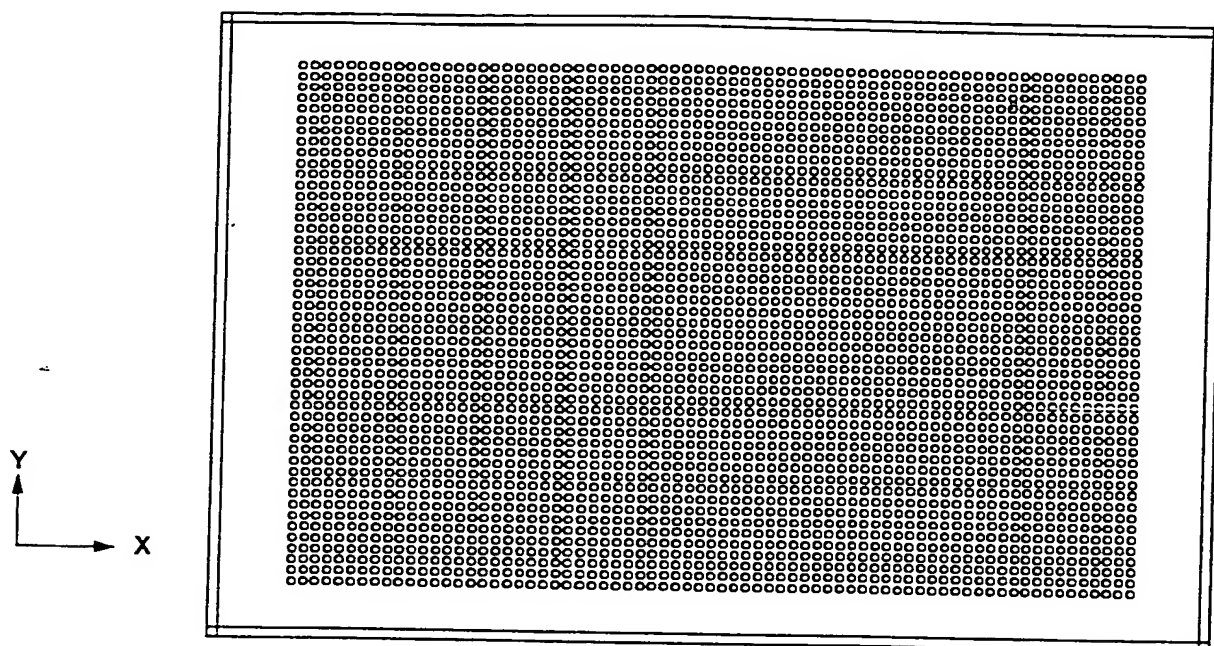
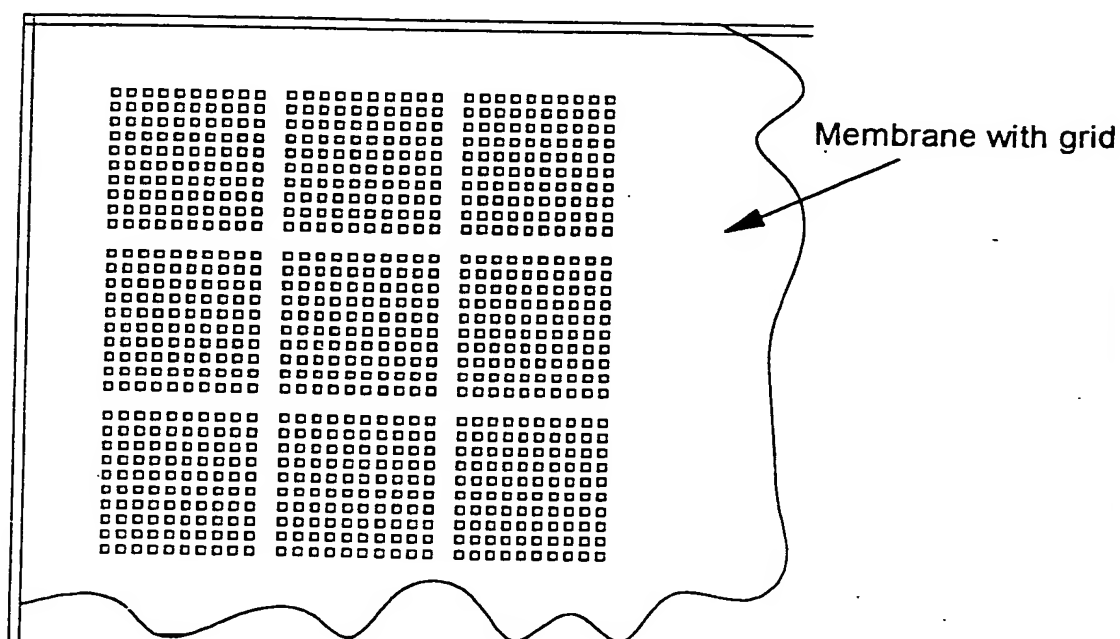


Fig 1(b). High density miniaturised well format presentation having 3456 wells.



Enlarged view of part of a high density gridded array of, eg, DNA

Fig 1(c).

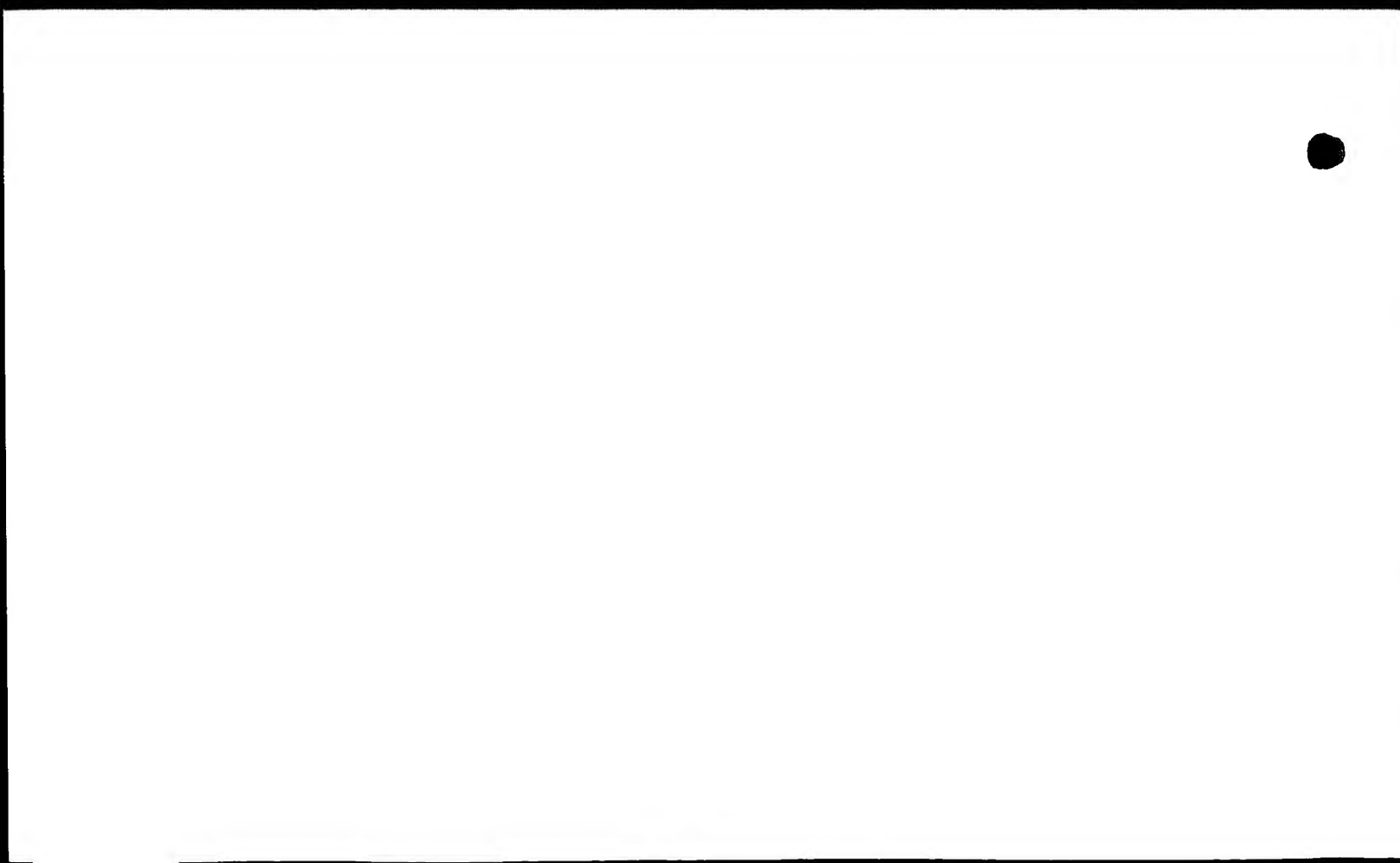
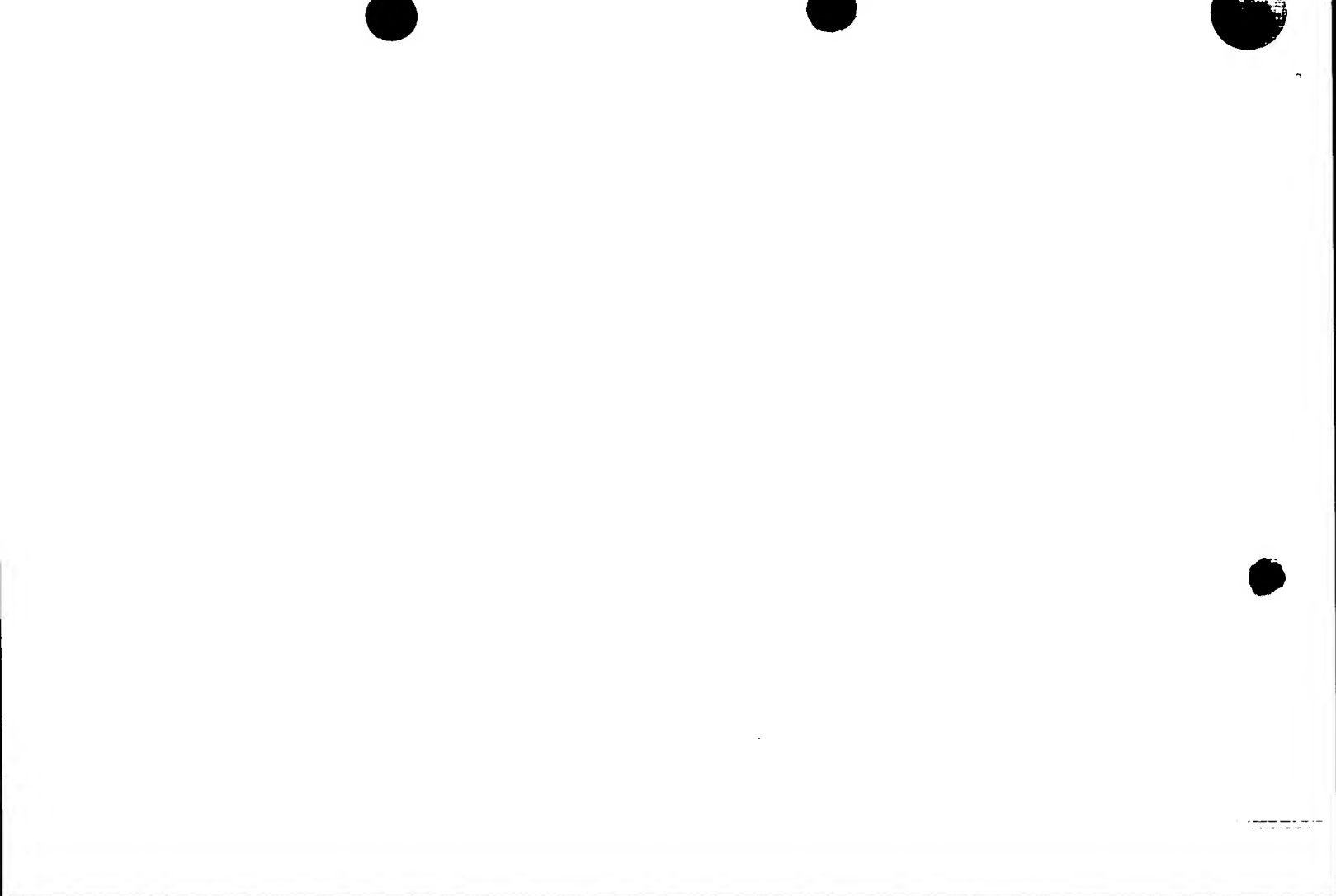
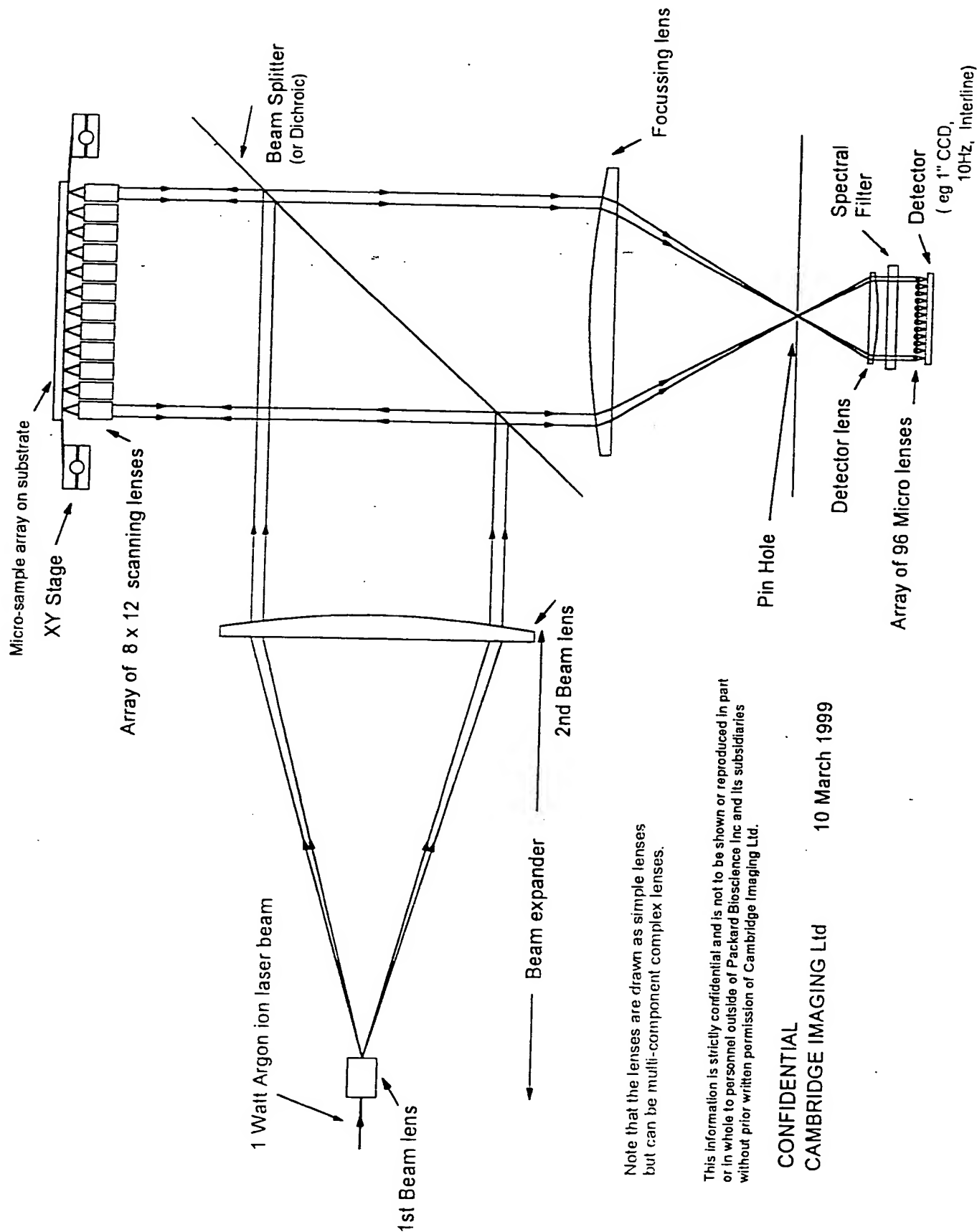


Fig 2 96 Channel Confocal Imager for measurement of Micro-sample Arrays



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10 March 1999

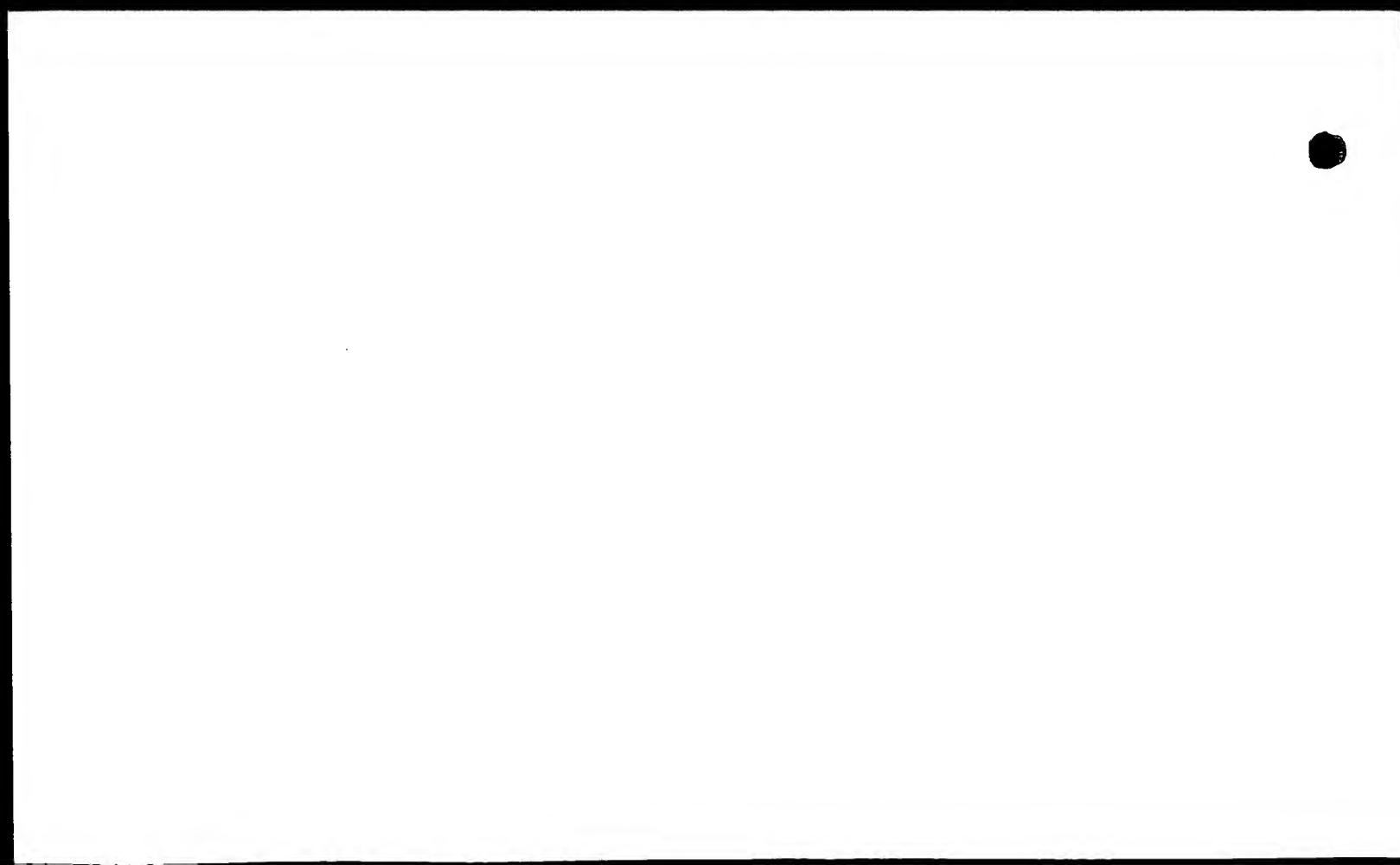
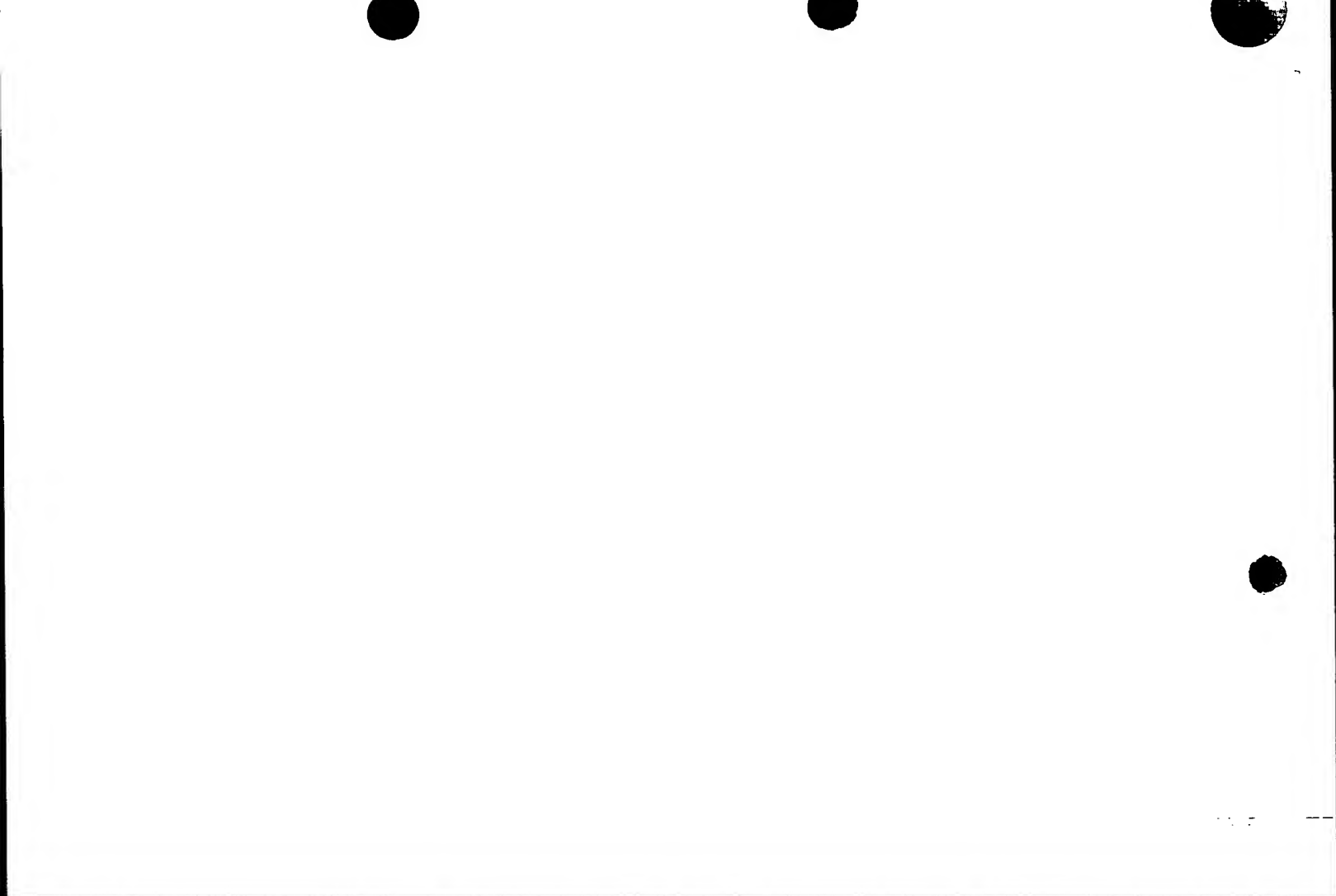
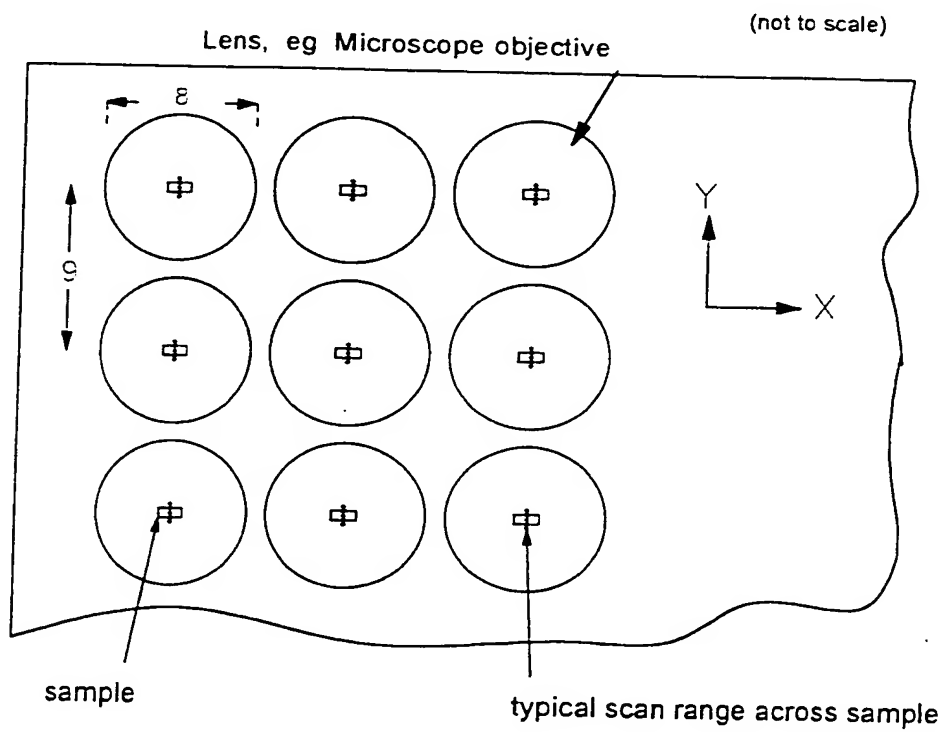


Fig 3



Plan view of scanning lenses over part of sample array

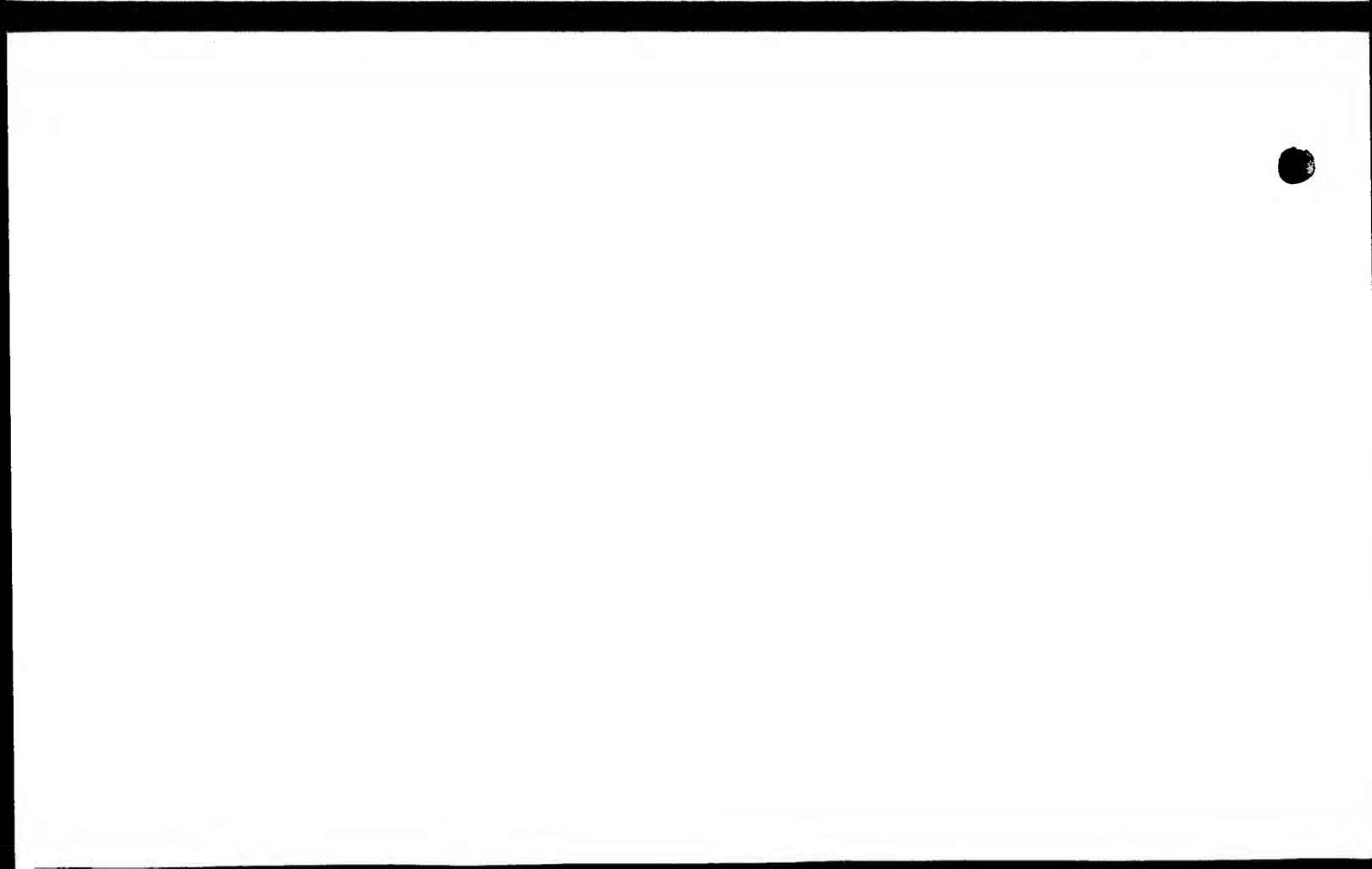
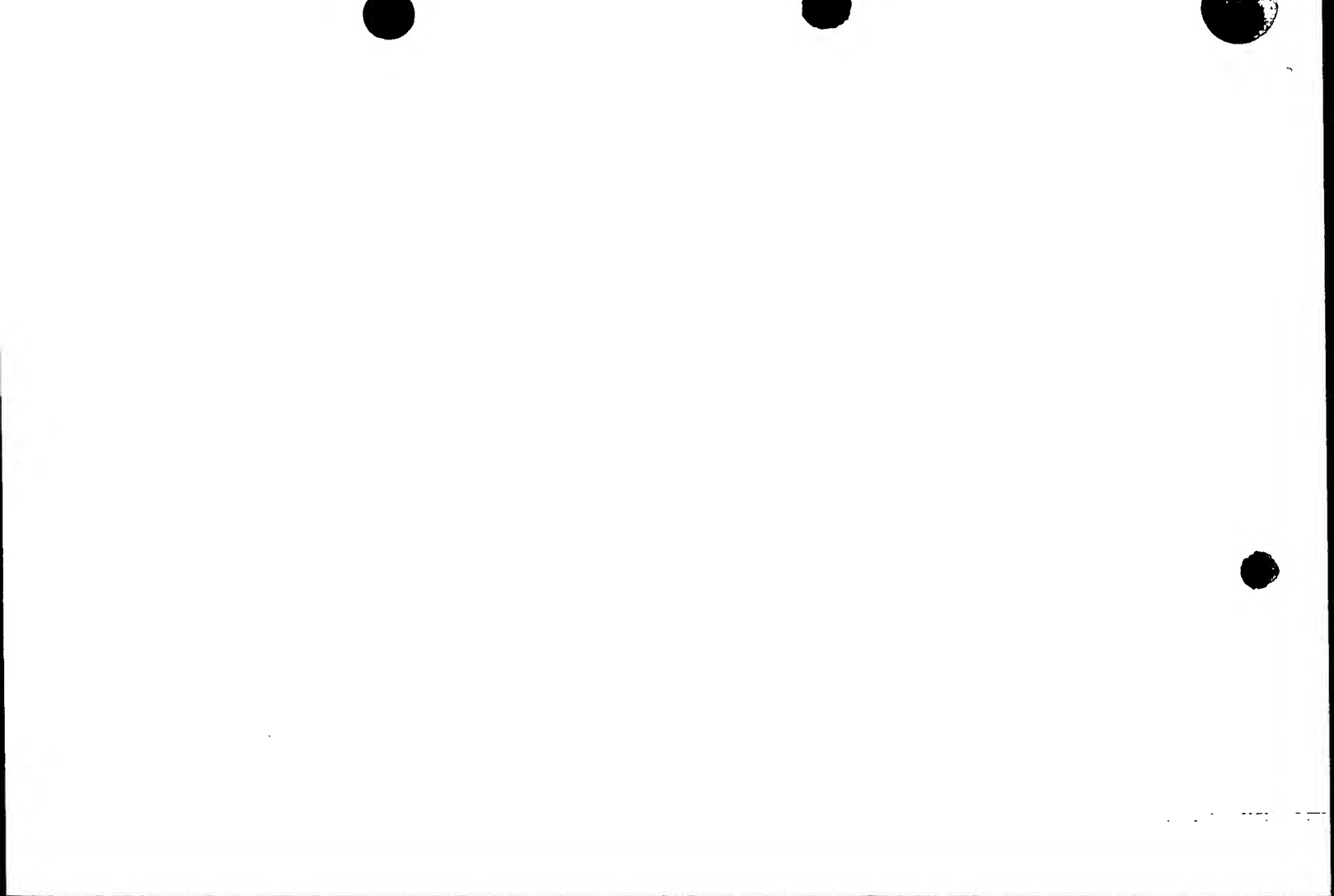
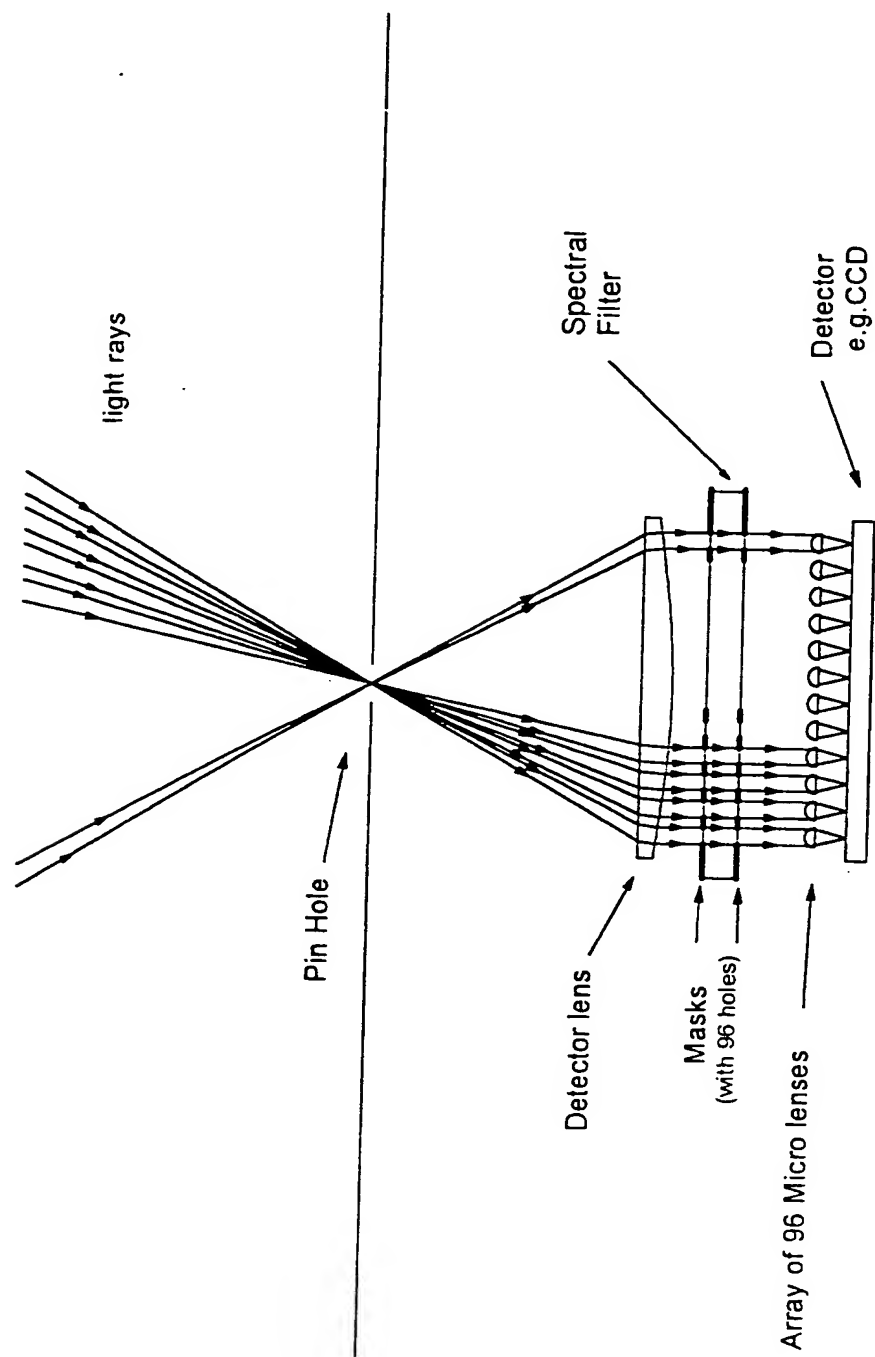


Fig 4. Detail of detector optical arrangement



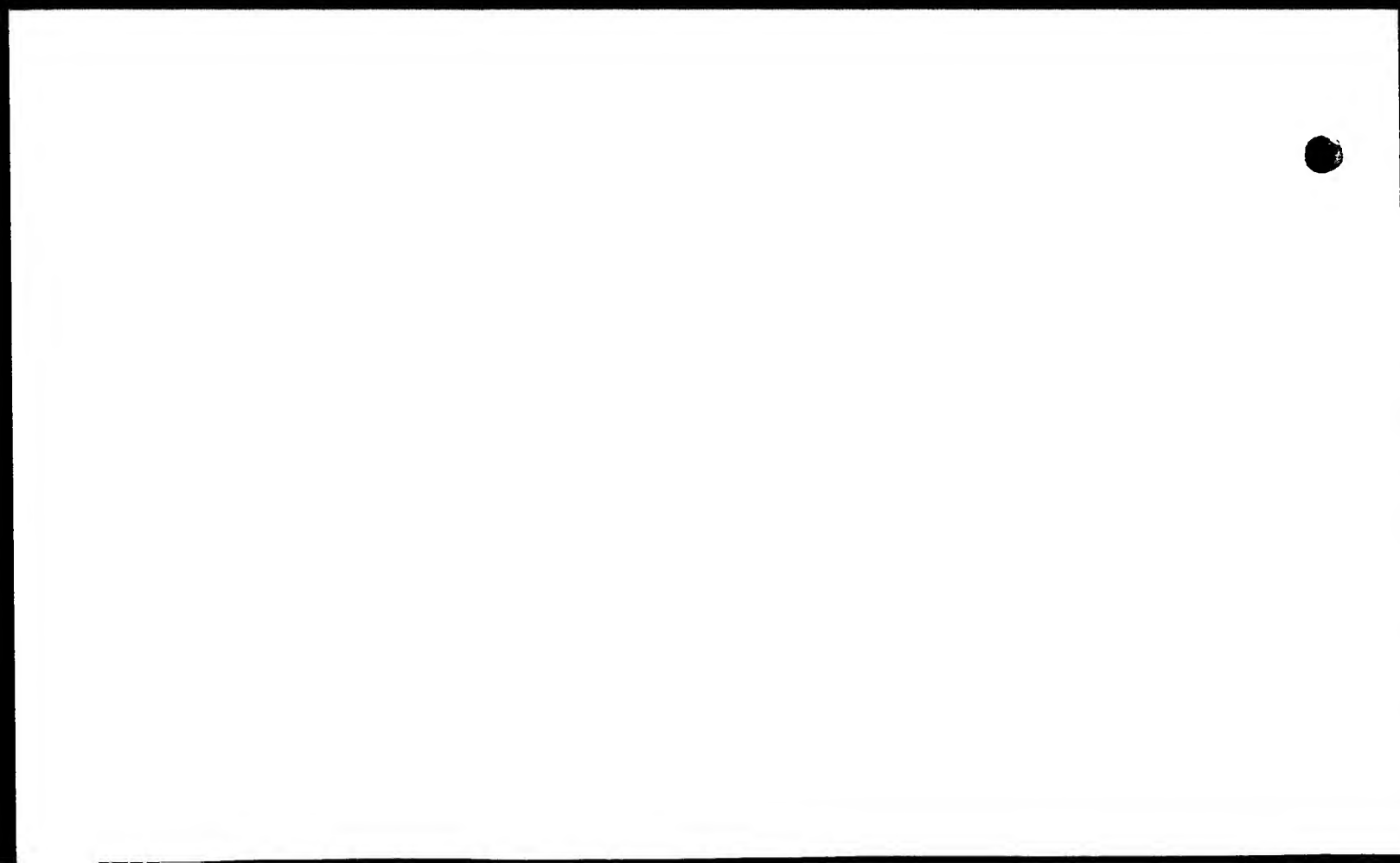
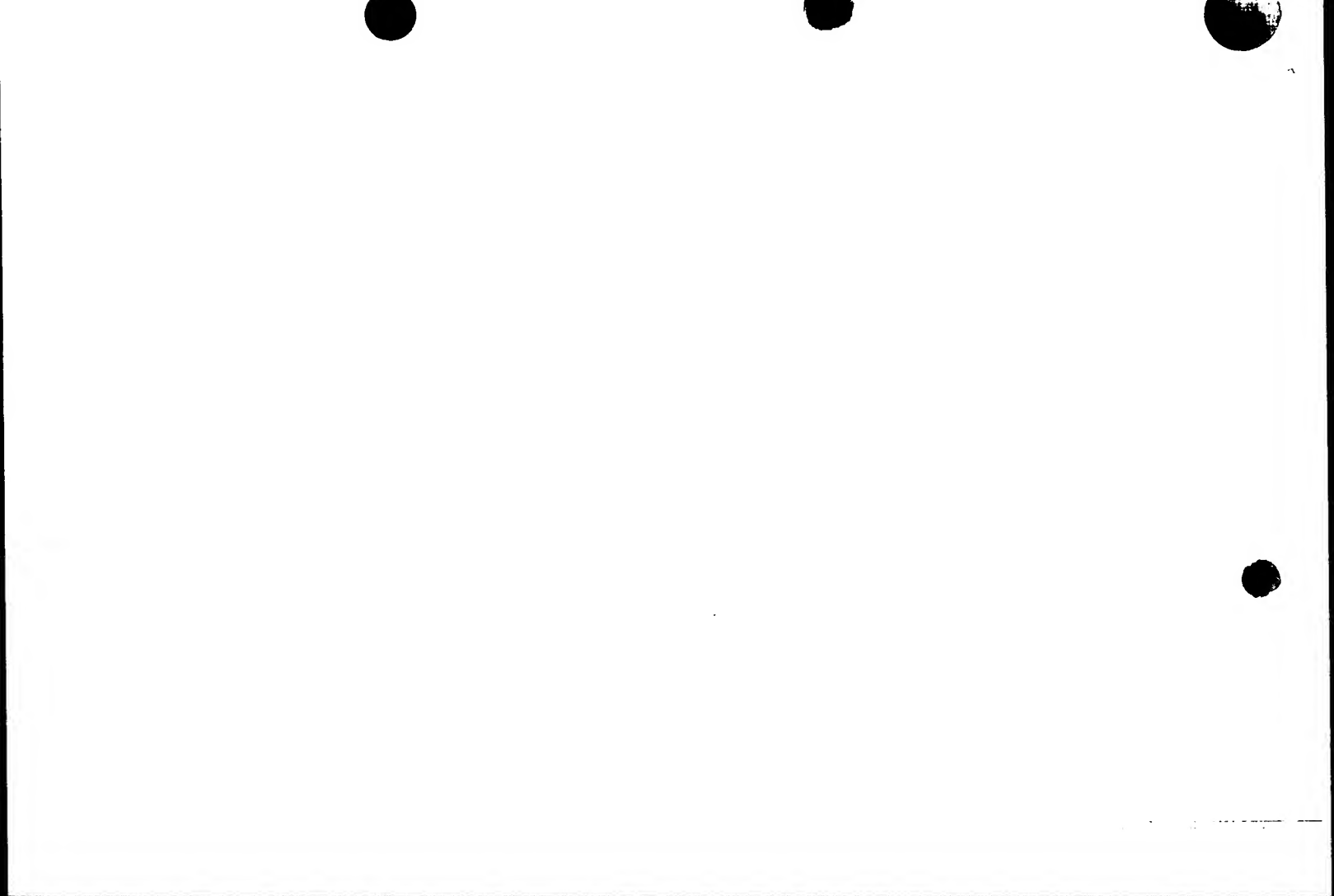
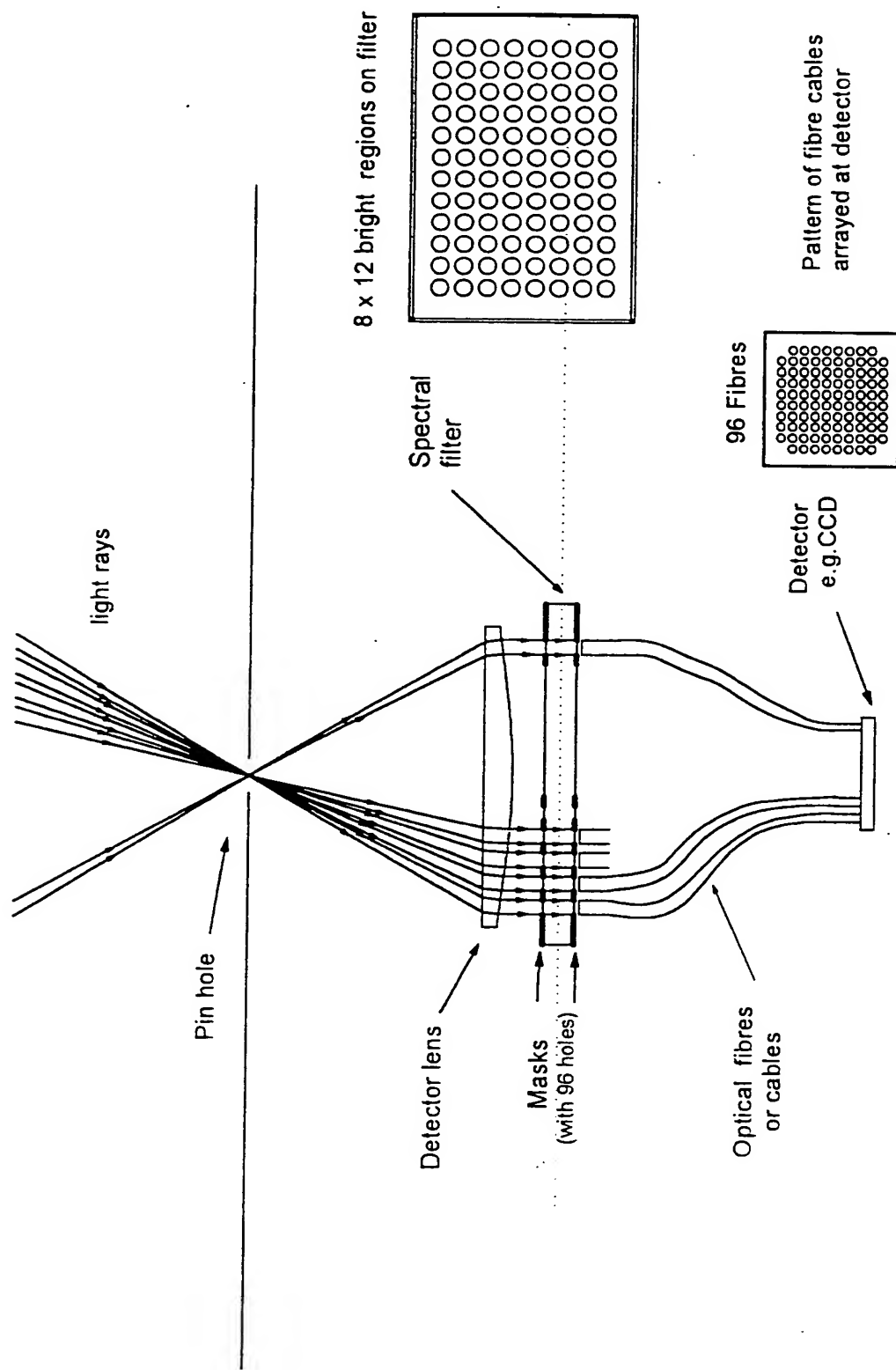


Fig 5. Detail of alternative detector optical arrangement



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